



Invasive Marine and Estuarine Animals of Hawai'i and other Pacific Islands

by Gary L. Ray

PURPOSE: New species of estuarine and marine animals are inadvertently or intentionally introduced into the waters of the United States every year (Figure 1). Variously referred to as introduced, nonindigenous (NIS), alien, nonnative, or exotic species, most pose little or no threat; however, a few have the potential to disrupt local ecosystems, fisheries, and human infrastructure. Such invasions directly impact the mission of the U.S. Army Corps of Engineers (USACE) through its responsibilities for construction and maintenance of harbors, ports, and waterways; erosion control; management of water resources; and wetland and coastal habitat restoration. The general biology and ecology of invasive estuarine and marine animals have been described in previous reports (e.g., Carlton (2001), Ray (2005)). This report is part of a series describing the biology and ecology of known invasive estuarine and marine animals in the major geographic regions of the United States. Invasive animals of the North Atlantic region are described and examples of species posing a specific threat to USACE activities are identified.

BACKGROUND: Invasive species are officially defined as “alien species whose introduction does or is likely to cause economic or environmental harm to human health” (Executive Order 13112, Federal Register (1999)). Any species removed from its native range has the potential to become invasive. Within a species’ normal range, predation, disease, parasites, competition, and other natural controls act to keep population levels in check (Torchin et al. 2003, Wolfe 2002). Once released from these controls, species abundances have the potential to reach levels that interfere with or displace local fauna. Such effects may occur immediately, after some period of delay, or may never be realized at all depending on the characteristics of the individual species and the conditions into which it is introduced.

Lists of estuarine and marine nonindigenous species are often dominated by mollusks, crustaceans, and polychaete worms; however, this may reflect their ease of identification and detection rather than the degree to which they are representative. Ultimately, it is an issue of an organism’s biological characteristics (e.g., reproductive capacity, growth rate, etc.) and not its taxonomic affinities that determine if it becomes invasive. Successful invaders tend to be those that are abundant over a large range in their native region, have broad feeding and habitat preferences, wide



Figure 1. Example of an invasive species, the Japanese Shore Crab, *Hemigrapsus sanguineus* (image courtesy of USGS)

physiological tolerances, short generation times, and high genetic variability (Erlich 1989, Williams and Meffe 1999). Despite the fact that we can identify these characteristics, predicting which species pose the greatest threat remains problematic. While many species possess these characters, most are not obvious in their native range, and the opportunity to be introduced and subsequent likelihood of survival are difficult to assess. The situation is further complicated by difficulty in distinguishing invaders from species with naturally wide distributions and those that are cryptogenic, that is, species whose original distributions are uncertain.

Predicting which habitats are likely to be invaded is much simpler. Invaded habitats tend to have low natural diversity, relatively simple (low-connectance) food webs, and a history of recent natural or anthropogenic disturbance (Williams and Meffe 1999). Estuaries and sheltered coastal areas are among the most invaded habitats, presumably due to the fact that they are naturally disturbed, low-diversity systems, and are historically centers of anthropogenic disturbance associated with navigation, industrial development, and urbanization.

Species are introduced by a variety of different mechanisms; however, most estuarine and marine species introductions are associated with shipping (Ruiz et al. 2000). Species capable of attaching to hard surfaces may be transported on ship hulls, navigational buoys, floatation devices, anchors, chains, ropes, and flotsam or jetsam (Carlton 2001). During the heyday of wooden-hulled ships, wood borers (e.g., shipworms) and species associated with "dry" ballast such as stones, rock, sand, or other materials were frequently introduced (Carlton and Hodder 1995). Presently, the largest single source of shipping-related introductions is ballast water (Carlton 1985, Lavoie et al. 1999). Ballast water is taken onboard vessels for a variety of purposes related to ship maneuverability and control (Carlton et al. 1995). Animals suspended in the water column or present in bottom sediments are taken in and then introduced to a new location when the ballast is pumped out.

Recently, concerns have also been raised regarding introductions of fish, invertebrates, and "live" rock from the aquarium trade (Padilla and Williams 2004, Weigle et al. 2005). The lionfish *Pterois volitans* may have been introduced when a private aquarium was demolished in the Miami area during Hurricane Andrew in 1992 (Hare and Whitfield 2003). Other introductions may result from accidental release of animals, inappropriate disposal of packing material by restaurants serving live seafood, and by the live bait industry. Many species have been deliberately introduced to develop new fisheries. For example, the Atlantic Striped Bass (*Morone saxatilis*) has been introduced both outside its normal geographic range and in non-native habitats (e.g., reservoirs) in much of the United States.

METHODS: As indicated in the official definition, invasive species are nonindigenous or alien to the region in which they are found. This does not mean that all NIS are invasive but rather that invasives constitute a subset of NIS. For the purposes of this report, the North Atlantic region is defined as covering the geographic area between the state of Maine and Chesapeake Bay. In order to create a list of known invasive species within this region, NISBase, a national database of NIS listings maintained by the Smithsonian Institution (<http://www.nisbase.org/nisbase/index.jsp>) was queried. Part of the National Exotic Marine and Estuarine Species Information System (NEMESIS), the database permits simultaneous searches of multiple NIS listings. Searches return up to 300 species and include links to individual species' fact sheets and collection data. Queries were

performed by state and included searches of the United States Geological Survey's Nuisance Aquatic Species (NAS) database (<http://nas.er.usgs.gov/>) and the Chesapeake Bay Exotic Marine and Estuarine Species Information System database. The resulting lists were amended as necessary after comparison with individual state NIS listings and other reports (Table 1), then edited to include only estuarine and marine species (Appendix A). Cryptogenic species were excluded from consideration due to the uncertainty of their origins. Only species known to be invasive or possessing the potential to become invasive are discussed.

Table 1
State NIS Listings and Other Sources Utilized in This Report

Benson and Richerson 2004
Benson et al. 2004
Carlton 2004
Massachusetts Aquatic Invasive Species Working Group 2002
Massachusetts Institute of Technology 2004
New York Sea Grant 2001
State of Vermont 2004

RESULTS: NIS listings for the North Atlantic region include 122 estuarine and marine animal species (Table 2). Not surprisingly, the greatest number of species was found among the molluscs (25 species) and fishes (19 species). Many of these represent species that were deliberately introduced, such as the oysters *Crassostrea gigas*, *C. ariakensis*, and *Ostrea edulis*, the Japanese littleneck clam *Venerupis philippinarum*, and fish including five species of west coast salmon

Table 2
Summary Results for Numbers of North Atlantic NIS by State

TAXON	ME	MA	NH	VE	RI	CN	NY	NJ	PA	DE	MD	VA	Total for Region
Protozoans		2						2			2	2	3
Sponges	1	2			1	1	1						2
Jellyfish											2	3	4
Anemones	1	2									1	2	3
Hydrozoans	2	1											2
Platyhelminths	1												1
Oligochaetes												1	1
Polychaetes	2				1						2	5	7
Bivalves	2	8			1	1	3	2			5	8	12
Gastropods	1	2			1			1		1	3	8	9
Nudibranchs	1	4										2	4
Copepods		1					1				2	2	3
Barnacles		3			1		1	1			2	2	5
Amphipods	2	3			1	1					1	6	10
Isopods	2	1						2			1	2	6
Mysids	1	1									1	1	2
Crabs	2	2	2		2	2	2	2		2	2	2	2
Insects	1	1									2	2	5
Bryozoans	6	7			1	1						1	8
Kamptozoans		1									2	2	3
Tunicates	4	11	3		5	4	1			1	1	9	12
Fish	4	8	3	7		4	11	4	14	4	9	10	19
Total	33	61	8	7	14	14	20	14	14	8	38	70	122

(*Onchorhynchus spp.*). A large number of indigenous fish are also included because they were stocked in so-called nonindigenous waters, i.e., waters where they don't naturally occur. These fish include all five species of shad and herring (*Alosa aestivalis*, *A. pseudoharengus*, *A. sapidissima*, *Dorosoma cepedianum*, and *D. petenense*), the mummichog *Fundulus heteroclitus*, the Atlantic cod *Gadus morhua*, the Atlantic salmon *Salmo salar*, and the striped bass *Morone saxatilis*. Several species may have been unintentionally introduced during shellfish introductions, including the oyster-disease-causing protozoans *Haplosporidium nelsoni* and *Perkinsus marinus* and the Japanese oyster drill *Eupleura sulcidentata*. The virus *Bonamia ostrea*, also fatal to oysters, is believed to have entered New England waters during shellfish introductions. Taken together, nearly 20 percent of all listed NIS in the North Atlantic are directly or indirectly attributable to deliberate introduction.

Fortunately, most of the fish introductions were of limited success and probably represent no direct threat to USACE operations. The unintentional introduction of the oyster diseases and predators, however, poses continuing difficulties to restoration of oyster habitat and stocks throughout the region.

An additional 37 NIS species (30 percent) are associated with hard structures and may contribute to fouling. Presumably introduced on ships' hulls or ballast water, they include two sponges (porifera), two hydrozoans, three anemones, five barnacles, eight bryozoans, three kamptozoans (entoprocts), twelve tunicates, and two polychaetes (Appendix A).

Distribution of NIS species among individual states varies widely, and to some degree, is a function of the amount of coastline and the intensity of efforts to detect NIS species (Table 1). Maine, Massachusetts, Maryland, and Virginia have extensive coastlines, a history of NIS monitoring, and the greatest numbers of recorded NIS species, while New Hampshire and Vermont with their relatively small coastlines have the fewest. For a review of factors potentially affecting the level of knowledge regarding NIS distributions and abundance, see Ray (2005).

DISCUSSION: Species most commonly identified as invasive or listed as of concern in North Atlantic states are the European green crab *Carcinus maenas*, the Japanese shore crab *Hemigrapsus sanguineus* (Figure 1), the white lace bryozoan *Membranipora membranacea*, the tunicates *Styella clavata* and *Botrylloides diagensis*, and the lionfish *Pterois volitans*. With the exception of the Japanese shore crab, the biology and potential impacts on USACE operations of these species have previously been described in Ray (2005).

Potential Threats to Infrastructure. The primary threat to infrastructure comes from several species of *Teredo* spp., wood-boring bivalve molluscs commonly called shipworms. Most shipworms, including the native species *Bankia gouldii*, are tolerant of a wide range of salinities and temperatures and low oxygen conditions. Shipworms are a potentially serious threat to wooden structures including boats, marinas, docks, and pilings. Cohen and Carlton (1995) report that a shipworm outbreak in San Francisco Bay resulted in \$615 million in damage during the 1920's. Likewise, severe damage was reported in Barnegat Bay, New Jersey and Long Island Sound, New York following outbreaks of *T. bartschi* associated with increased water temperatures and salinities in power station effluents (Hoagland 1983). These pests can be effectively controlled by chemical treatment (e.g., creosote) of wood or use of alternative materials (Highley 1999).

Because of their ability to foul coastal structures such as intake pipes, a variety of introduced fouling species (e.g., sponges, barnacles) have the potential to interfere with USACE operations. Thus far, only the hydroids *Cordylophora caspia* and *Garveia franciscana* have been positively identified as posing such a threat (Massachusetts Institute of Technology 2004). The tunicate *Didemnum lahille* has recently interfered indirectly with a navigation project. This species is undergoing a major outbreak in New England (National Marine Fisheries Northeast Science Center 2004) and resource agencies have voiced the concern that dredging operations might contribute to its spread.¹

Potential Threats to Habitat Restoration.

Green crabs feed voraciously on shellfish and may disrupt attempts to restore softshell clam (*Mya arenaria*) stocks. Another species that may affect shellfish restoration is the whelk *Rapana venosa* (Figure 2). Presently limited to Chesapeake Bay, it is likely to expand its range to include most of the North Atlantic and

portions of the South Atlantic (Mann and Harding 2003). This species is a voracious predator of shellfish, including hard clams (*Mercenaria mercenaria*), softshell clams, and oysters. Control efforts currently include offering a bounty for whelks and encouraging local restaurants to develop recipes incorporating this species.



Figure 2. The Veined Rapa Whelk, *Rapana venosa* (image courtesy of the Jacksonville Florida Shell Club)

Japanese shore crab. The Japanese shore crab is a native of the western Pacific Ocean characteristically found in low intertidal and subtidal zones with rock or cobble substrates. First detected in New Jersey in 1988 (McDermott 1991), it has since spread north into southern New England (Lohrer and Whitlach 1997) and south into Chesapeake Bay. It has quickly become one of the most abundant crabs in the rocky intertidal zone, displacing native xanthid crabs and even the invasive green crab from areas where their distributions overlap (Lohrer and Whitlach 1997, Jensen et al. 2002). It has an extended breeding season and tolerates salinities as low as 15 ppt as larvae (Epifaunio et al. 1998). Adults appear to prefer salinities of 20 ppt or higher. The Japanese shore crab has broad food preferences including macroalgae, snails, mussels, and barnacles (Lohrer and Whitlach 1997; Bourdeau and O'Connor 1999, Percival and Wilson 2001). While it seems doubtful this species will directly interfere with USACE operations, the issue of incidentally contributing to its spread has arisen in situations where riprap or other coarse materials are being placed in the intertidal zone². Since there are presently no known control methods, the issue remains problematic.

¹ Personal communication, 2004, Catherine Rogers, Ecologist, U.S. Army Engineer District, New England, Concord, MA.

² Personal communication, 2004, Robert Will, U.S. Army Engineer District, New York.

European periwinkle. An additional species of concern in northern New England is the common or European periwinkle *Littorina littorea*. Introduced sometime in the mid-19th century, it has spread throughout the North Atlantic region. It has displaced native mud snails *Nassarius obsoletus*, also known as *Ilyanassa obsoleta*, from many of their habitats and appears to compete with native *Littorina* species for food (Brenchley and Carlton 1983, Yamada and Mansour 1987). Bertness (1984) has shown that grazing and “bulldozing” of sediments by this species prevents accumulation of fine sediments and development of an algal canopy on rocky intertidal sites. When experimentally removed, the snail-free sites gradually became areas of soft sediment. Likewise, removal of snails resulted in increased growth of the salt marsh cord grass *Spartina alterniflora* due to a decrease in herbivory by the snail and an increase in sediment accumulation, which promotes expansion of the root mat.

Veined rapa whelk. *Rapana venosa*, the veined rapa whelk, a native of the Sea of Japan, was first detected in Chesapeake Bay in 1998 (Mann and Harding 2000). Already known as an invasive species in the Black, Adriatic, and Aegean Seas, it is thought to have entered the United States via ballast water. It prefers hard sandy bottoms with salinities of 18-28 ppt and can reach a total length of 150 mm. While juveniles feed on barnacles, mussels, and newly settled and very young oysters, adults feed heavily on soft clams and hard clams. This species has the potential to affect restoration of shellfish habitat in Chesapeake Bay and elsewhere. Distribution of this species is presently limited to portions of Chesapeake Bay, but it will likely spread north to Cape Cod and south to Cape Hatteras (Mann and Harding 2003).

***M. membracea*.** The European bryozoan *M. membracea* is believed to have first arrived in U.S. waters sometime in the 1980's (Berman et al. 1990). Now the dominant fouling animal of laminarian kelp in the North Atlantic, it has also been collected on the Pacific coast (Berman et al. 1992, Lambert et al. 1992). Colonies of this animal can become so abundant that the kelp blades are literally weighed down, making the blades more susceptible to damage by wave action (Dixon et al. 1981). Such impacts can directly interfere with attempts to restore kelp habitat.

Potential Threats to Human Health. The lionfish *Pterois volitans*, a native of the Indian Ocean and western Pacific coral reefs, is a favorite aquarium fish because of its colorful appearance. It bears poisonous dorsal, anal, and pelvic spines. The toxin is painful, but is generally not fatal. The lionfish was first noticed in Florida waters in 1994 and may have been introduced when a private aquarium was demolished in the Miami area during Hurricane Andrew in 1992 (Hare and Whitfield 2003). Associated with hard bottoms, coral reefs, and artificial substrates from 26 to 79 m deep, it is generally found on ledges and crevices from Florida to Cape Hatteras, North Carolina. It has also been reported in Puerto Rico. Lionfish feed mostly upon small fish, crabs, and shrimp; however, their impact on local ecosystems has not been assessed.

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NOTE: The contents of this technical note are not to be used for advertising, publication, or promotional purposes. Citation of trade names does not constitute an official endorsement or approval of the use of such products.

APPENDIX A: NIS LISTINGS BY STATE FOR THE NORTH ATLANTIC REGION

Species	Group	ME	MA	NH	VE	RI	CN	NY	NJ	PA	DE	MD	VA
<i>Perkinsia marinus</i>	Protozoa		MA						NJ			MD	VA
<i>Haplosporidium nelsoni</i>	Protozoa		MA						NJ			MD	VA
<i>Bonamia ostrea</i>	Virus	ME	MA										
<i>Cliona vastifica</i>	Porifera		MA										
<i>Halichondria bowerbankia</i>	Porifera	ME	MA			RI	CN	NY					
<i>Blackfordia virginica</i>	Hydromedusa											MD	VA
<i>Moerisia lysonsi</i>	Hydromedusa											MD	
<i>Maeotias inexpectata</i>	Hydromedusa												VA
<i>Maeotias marginata</i>	Hydromedusa												VA
<i>Nematostella vectensis</i>	Anemone											MD	VA
<i>Diadumene lineata</i>	Anemone	ME	MA										VA
<i>Sagartia elegans</i>	Anemone		MA										
<i>Cordylophora caspia</i>	Hydrozoa	ME											
<i>Convoluta convoluta</i>	Platyhelminth	ME											
<i>Tubificoides heterochaetus</i>	Oligochaete												VA
<i>Demonax flectatus</i>	Polychaete												VA
<i>Demonax sp.</i>	Polychaete												VA
<i>Namanereis littoralis</i>	Polychaete												VA
<i>Boccardiella ligerica</i>	Polychaete											MD	VA
<i>Boccardiella proboscidea</i>	Polychaete	ME											
<i>Ficopomatomus enigmaticus</i>	Polychaete											MD	VA
<i>Janua pagenstecheri</i>	Polychaete	ME				RI							
<i>Bankia gouldii</i>	Bivalve											MD	
<i>Crassostrea ariakensis</i>	Bivalve												VA
<i>Crassostrea gigas</i>	Bivalve		MA									MD	VA
<i>Cyrenoida floridana</i>	Bivalve											MD	VA
<i>Ischadium recurvum</i>	Bivalve											MD	VA
<i>Ostrea edulis</i>	Bivalve	ME	MA			RI							
<i>Mytilopsis leucopaeata</i>	Bivalve		MA					NY					
<i>Rangia cuneata</i>	Bivalve		MA					NY				MD	VA
<i>Teredo bartschi</i>	Bivalve		MA				CN		NJ				VA
<i>Teredo furcifera</i>	Bivalve		MA						NJ				VA
<i>Teredo navalis</i>	Bivalve	ME	MA					NY					VA
<i>Venerupis philippinarum</i>	Bivalve		MA										
<i>Assiminea succinea</i>	Gastropod											MD	VA
<i>Eupleura sulcidentata</i>	Gastropod												VA
<i>Littorina littorea</i>	Gastropod	ME	MA			RI			NJ	DE			VA
<i>Melampus floridanus</i>	Gastropod											MD	VA
<i>Mysotella myosotis</i>	Gastropod		MA										

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Species	Group	ME	MA	NH	VE	RI	CN	NY	NJ	PA	DE	MD	VA
<i>Pyostella myosotis</i>	Gastropod												VA
<i>Rapana venosa</i>	Gastropod												VA
<i>Stramonita haemastoma</i>	Gastropod											MD	VA
<i>Vitrinella floridana</i>	Gastropod												VA
<i>Tritonia plebeia</i>	Nudibranch	ME	MA										
<i>Placida dendritica</i>	Nudibranch		MA										VA
<i>Tenellia adspersa</i>	Nudibranch		MA										VA
<i>Argulus japonicus</i>	Copepod										MD	VA	
<i>Eurytemora affinis</i>	Copepod		MA					NY					VA
<i>Mytilicola</i> sp.	Copepod										MD		
<i>Balanus amphitrite</i>	Barnacle		MA					NY	NJ		MD	VA	
<i>Balanus subalbidus</i>	Barnacle		MA										
<i>Chthamalus fragilis</i>	Barnacle					RI							
<i>Elminius modestus</i>	Barnacle		MA										
<i>Loxothylacus panopaei</i>	Barnacle										MD	VA	
<i>Amphilochus</i> sp.	Amphipod											VA	
<i>Apocorophium acutum</i>	Amphipod											VA	
<i>Monocorophium acheruscium</i>	Amphipod											VA	
<i>Monocorophium insidiosum</i>	Amphipod											VA	
<i>Corophium volutator</i>	Amphipod	ME	MA										
<i>Caprella mutica</i>	Amphipod	ME											
<i>Cheurla terebrans</i>	Amphipod		MA										
<i>Gitanopsis</i> sp.	Amphipod										MD	VA	
<i>Microdeutops gryllotalpa</i>	Amphipod		MA	RI			CN						
<i>Stenothoe gallensis</i>	Amphipod											VA	
<i>Ligia exotica</i>	Isopod								NJ		MD	VA	
<i>Ligia oceanica</i>	Isopod	ME											
<i>Ianiropsis</i> sp.	Isopod	ME											
<i>Limnoria</i> sp.	Isopod		MA										
<i>Sphaeroma terebrans</i>	Isopod											VA	
<i>Synidotea laevoidorsalis</i>	Isopod								NJ				
<i>Americamysis almyra</i>	Mysid										MD	VA	
<i>Praunus flexuosus</i>	Mysid	ME	MA										
<i>Carcinus maenas</i>	Crab	ME	MA	NH		RI	CN	NY	NJ		DE	MD	VA
<i>Hemigrapsus sanguineus</i>	Crab	ME	MA	NH		RI	CN	NY	NJ		DE	MD	VA
<i>Anisolabis maritima</i>	Insect	ME	MA										VA
<i>Brachydeutera longipes</i>	Insect										MD		
<i>Placopsidella erandis</i>	Insect										MD		
<i>Placopsidella grandis</i>	Insect											VA	
<i>Procanace diannae</i>	Insect												
<i>Aetea anguina</i>	Bryozoan		MA										
<i>Membranipora membranacea</i>	Bryozoan	ME	MA			RI	CN						
<i>Stephanella hina</i>	Bryozoan		MA										

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Species	Group	ME	MA	NH	VE	RI	CN	NY	NJ	PA	DE	MD	VA
<i>Alyconidium</i> sp.	Bryozoan	ME											
<i>Bugula neritina</i>	Bryozoan	ME	MA										VA
<i>Bulbella abscondita</i>	Bryozoan		MA										
<i>Tangenella muelleri</i>	Bryozoan		MA										
<i>Tangenella appendiculata</i>	Bryozoan		MA										
<i>Barentsia benedini</i>	Kamptozoan		MA										VA
<i>Barentsia carteri</i>	Kamptozoan											MD	
<i>Loxosomatoides laevis</i>	Kamptozoan										MD	VA	
<i>Ascidia aspersa</i>	Tunicate		MA			RI	CN						VA
<i>Botrylloides diegensis</i>	Tunicate	ME	MA	NH		RI	CN						
<i>Botrylloides violaceus</i>	Tunicate	ME	MA										VA
<i>Botryllus schlosseri</i>	Tunicate	ME	MA										VA
<i>Didemnum cf. lahillei</i>	Tunicate	ME	MA	NH		RI							
<i>Diplosoma macdonaldi</i>	Tunicate	ME	MA			RI	CN						
<i>Diplosoma listerianum</i>	Tunicate		MA										VA
<i>Ecteinascidia turniata</i>	Tunicate												VA
<i>Mogula manhatiensis</i>	Tunicate	ME	MA								NJ*	MD	VA
<i>Styela clavata</i>	Tunicate	ME	MA	NH		RI	CN	NY					VA
<i>Styela canopus</i>	Tunicate		MA										VA
<i>Styela plicata</i>	Tunicate		MA										VA
<i>Alosa aestivalis</i>	Fish			VE				NY		PA			VA
<i>Alosa pseudoharengus</i>	Fish		MA	NH	VE			NY		PA		MD	VA
<i>Alosa sapidissima</i>	Fish		MA		VE			NY		PA		MD	VA
<i>Anguilla rostrata</i>	Fish									PA			
<i>Dorosoma cepedianum</i>	Fish			VE				NY		PA			VA
<i>Dorosoma petenense</i>	Fish									PA	DE	MD	VA
<i>Fundulus heteroclitus</i>	Fish			NH						PA			
<i>Gadus morhua</i>	Fish											MD	VA
<i>Gobionellus hastatus</i>	Fish		MA										
<i>Hypsoblennius ionthas</i>	Fish		MA										
<i>Morone saxatilis</i>	Fish								NJ	PA		MD	VA
<i>Oncorhynchus gorbuscha</i>	Fish							NY		PA		MD	
<i>Oncorhynchus keta</i>	Fish	ME											
<i>Oncorhynchus kisutch</i>	Fish	ME	MA				CN	NY		PA	DE	MD	VA
<i>Oncorhynchus nerka nerka</i>	Fish		MA	VE		CN	NY		PA				
<i>Oncorhynchus tshawytscha</i>	Fish	ME	MA	NH	VE	CN	NY	NJ	PA	DE	MD	VA	
<i>Petromyzon marinus</i>	Fish							NY		PA			
<i>Pterois volitans</i>	Fish							NY	NJ				
<i>Salmo salar</i>	Fish	ME	MA		VE	CN	NY	NJ	PA	DE	MD	VA	

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